

Evidence for Linkage of a Candidate Chromosome 1 Region to Human Systemic Lupus Erythematosus

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Abstract

Genetic susceptibility confers significant risk for systemic lupus erythematosus (SLE). The MHC region and other polymorphic loci have been associated with SLE. Because more compelling evidence for an involvement of a genetic locus includes linkage, we tested a candidate region homologous to a murine SLE susceptibility region in 52 SLE-affected sibpairs from three ethnic groups. We analyzed seven microsatellite markers from the human chromosome 1q31–q42 region corresponding to the telomeric end of mouse chromosome 1, the region where specific manifestations of murine lupus, including glomerulonephritis and IgG antichromatin, have been mapped. Comparing the mean allele sharing in affected sibpairs of each of these seven markers to their expected values of 0.50, only the five markers located at 1q41–q42 showed evidence for linkage ($P = 0.0005$ – 0.08). Serum levels of IgG antichromatin also showed evidence for linkage to two of these five markers ($P = 0.04$), suggesting that this phenotype is conserved between mice and humans. Compared to the expected random distribution, the trend of increased sharing of haplotypes was observed in affected sibpairs from three ethnic groups ($P < 0.01$). We concluded that this candidate 1q41–q42 region probably contains a susceptibility gene(s) that confers risk for SLE in multiple ethnic groups. (*J. Clin. Invest.* 1997; 99:725–731.) Key words: human SLE • affected sibpairs • susceptibility • linkage analysis • homology

Introduction

Systemic lupus erythematosus (SLE)¹ is an autoimmune rheumatic disease characterized by the tissue disposition of autoan-

tibodies and immune complexes leading to tissue injury (1). The serological hallmark of this disease is elevated serum levels of IgG antibodies to nuclear constituents such as double-stranded DNA (dsDNA) and chromatin. Among these autoantibodies, IgG anti-dsDNA antibodies play a major role in the development of lupus glomerulonephritis (GN) (2–3). Mechanisms by which these autoantibodies are induced remain unclear. Chromatin and/or nucleosomes which are released by apoptotic cells in SLE may be autoantigens that induce autoimmune responses, including antibodies to dsDNA (4–6). The actual mechanisms may be best delineated after the susceptibility genes for SLE are described, because it appears that genetic factors exert the greatest influence on autoantibody production and on predisposition to SLE (7).

Studies of populations, segregation of disease in families, and twin concordance rates have consistently demonstrated the importance of genetic influences on SLE (7). The risk for siblings of SLE patients to develop disease is estimated to be 20 times higher than that of the general population (7–9). The genetic basis for SLE is complex, with an unknown but non-Mendelian mode of inheritance. Many investigators have reported that certain MHC class II alleles or complement gene deficiencies (homozygous deficiency of C1q, C2, and C4) are associated with SLE in most ethnic groups studied (7, 10, 11). Other alleles at polymorphic genes have also been reported to be associated with SLE, but their contributions are now well defined (reviewed in 7). These polymorphic candidate genes include those that encode mannose-binding protein, Ro/SSA, CR1, IL-6, Ig Gm and Km allotypes, TcR, TNF α , Fc γ RIIA, and Hsp-70 (12–24). Some of these candidate genes may confer risk to subsets of SLE patients. For example, the Fc γ RIIA polymorphism (the gene encoding a 40-kD Fc γ R expressed on human mononuclear phagocytes and neutrophils) has been associated with SLE GN in African Americans but not in Caucasians in one study (22), and in Caucasians in another study (23). A possible genetic linkage of Fc γ RIIA with lupus has been reported in a sample of four African-American multiplex families (25). The Fc γ RIIA and Fc γ RIIIA genes are located at the chromosome 1q23–q24 region.

Elements complicating the study of disease-causing genes in genetically complex diseases such as human SLE include ethnic diversity, clinical heterogeneity (and presumably genetic heterogeneity), reduced penetrance, and the effect of environment (26, 27). In contrast, murine models of spontaneous lupus in inbred strains are less complex. Genetic studies of murine SLE have identified susceptibility loci in several inbred strains which spontaneously develop SLE GN (28). These studies have included genome-wide searches for evidence of linkage using backcrosses or F2 intercrosses of lupus mice such

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1. Abbreviations used in this paper: dsDNA, double-stranded DNA; GN, glomerulonephritis; IDDM, insulin-dependent diabetes mellitus; SLE, systemic lupus erythematosus.

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as MRL/lpr, NZB/NZW, and NZM/Aeg2410 (29–33). The distal end of mouse chromosome 1 was shown to predispose to specific manifestations of SLE, including GN, IgG antibodies to chromatin, DNA and histone, and splenomegaly (30, 31, 33–36). Because important susceptibility genes may be conserved between mice and humans, we have chosen to focus initially on the chromosome 1q21–q42 region corresponding to the murine SLE susceptibility region identified in three independent crosses (30, 31, 33). Here we report that the tested markers present in the human chromosome 1q41–q42 region show evidence of linkage to human SLE, as well as to the presence of IgG antichromatin autoantibodies.

Methods

Family enrollment. 43 families with at least two siblings who met the revised 1982 American College of Rheumatology classification criteria for SLE (37) were recruited. These families contained 52 affected sibpairs; 40 families contained 2 affected sibs (e.e., 40 affected sibpairs), 2 contained 3 affected sibs (for a total of 6 affected sibpairs) and one contained 4 affected sibs (6 affected sibpairs total). 30 families were from Southern California, 8 from Taiwan, 2 from England, 2 from China, and 1 from Korea. Both parents and unaffected sibs, if available, were included in the study to enable the genetic markers to be maximally informative for identity-by-descent status to be used in the linkage tests. 19 sibpair families had both parent available, 12 had one parent, and the remaining 12 had no available parent for testing. The SLE patients ranged in age from 10 to 75 yr, sex distribution was 74% females and 26% males. Siblings of SLE patients were classified as unaffected by self-reporting. The youngest unaffected sib in our cohort was 14 yr old. Because of the wide range of age of onset for SLE (< 1 to over 60 yr old; 9), no age limit for unaffected sibs can be effectively used to exclude those who may develop SLE at a later time. Each participant donated peripheral blood for serum and DNA. This study was approved by the Human Subject Protection Committee of the University of California, Los Angeles (UCLA).

Clinical studies. Data from medical records for each SLE patient/proband were reviewed by UCLA rheumatologists and entered into the SLE database. This database was used for confirmation of diagnosis of SLE (requiring at least four of the 11 American College of Rheumatology criteria; 37). DNA and serum were stored for each proband and family member.

Antichromatin IgG antibodies. Histone H1-stripped chromatin (referred to hereafter as chromatin) was prepared from calf thymus

as described as Burlingame and Rubin (39). Chromatin (10 µg/ml native DNA) was diluted in PBS and applied in 50-µl aliquots to Immulon II plates as antigen for 1 h at room temperature. After washing, these wells were blocked with 100 µl of 10% FCS plus 0.05% Tween 20 in PBS for 1 h. All serum samples were diluted 1:100 in 5% FCS in PBS. 50 µl of diluted serum samples were applied to the wells in duplicates. After washing with 0.1% Tween 20 in PBS, goat anti-human IgG (Fc-specific) conjugated with alkaline phosphatase was applied. Subsequently, these plates were washed and color reaction was developed 30 min after the addition of substrate.

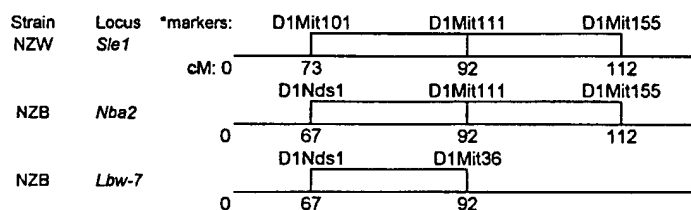
Anti-dsDNA IgG antibodies. This ELISA was performed as previously described (39) using serum samples diluted 1:100 in 5% FCS in PBS. Each plate contained known positive and negative controls in addition to test serum samples. The DNA used was calf thymus dsDNA treated to remove single strand nicks. Because the levels of IgG anti-dsDNA fluctuate with time in SLE patients, a single serum sample from studied subjects may provide false negative results. However, historical information regarding anti-DNA titers extracted from medical records of SLE patients could not be used for quantitative trait analysis because of (a) missing data from some patients, (b) absence of normal ranges from some laboratory reports, and (c) use of multiple methods, none of which is standardized.

Genotyping. DNA was isolated from peripheral blood cells. Microsatellite markers at or near the specific candidate chromosomal region were typed. The primers for these markers were purchased from Research Genetics (Huntsville, AL). Microsatellite genotyping was determined by scoring the size of PCR products. PCR was performed in a 96-well plate using a thermocycler (MJ Research, Inc., Watertown, MA) which was programmed for 95°C for 1 min, 58°C for 30 s, and 72°C for 1 min, with 25 cycles. The labeled PCR products were denatured and separated on a 5% sequencing gel.

The candidate region. Murine susceptibility regions on chromosome 1, their syntenic human chromosomal regions, and the approximate location of seven tested microsatellite markers are depicted in Fig. 1. These seven markers, located on 1q31–q42 spanning an ~27-cM region, were used to genotype the SLE multiplex families. Both the genetic distance and the cytogenetic assignment were based on the composite map, which integrates physical, radiation hybrid, genetic linkage, and cytogenetic assignment (40). The sex-averaged distance, chromosomal location and heterozygosity index of each marker are shown in Table I.

Analytic strategy including statistical linkage analyses. In order to assess cosegregation of the lupus phenotype with markers in the region of interest on chromosome 1, we employed several nonparametric linkage tests, i.e., tests for which the model of inheritance of the disease does not have to be hypothesized (41). Each of the seven

A. Murine Susceptibility Regions



B. Human Homologous Region

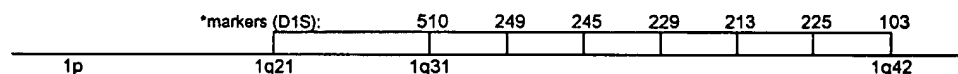


Figure 1. Murine susceptibility regions (A), their human homologous region (B), and relative positions of tested markers. (A) Murine SLE susceptibility regions are located within the distal end of mouse chromosome 1 marked by *D1Nds1* and *D1Mit155*, which correspond to the human chromosome 1q21–q42 region as shown in (B). The seven markers tested in this study are located within chromosome 1q31–q42 region.

* Approximate marker position (not to scale) was

obtained from the mouse genome database at <http://www.informatics.JAX.org/>. The mouse/human homology was obtained at <http://www3.ncbi.nlm.gov/Homology/>. The human chromosome 1 composite map was obtained at <http://cedar/genetics/soton.ac.uk/public-html/>.

Table I. Markers Tested for Linkage to SLE and SLE-related Traits

Marker No.	Marker locus	5' Primer	3' Primer	Chromosome locus*	Genetic distance (cM) [†]	HET [‡]
1	DIS510	AACCCGAGGTGTCTGTGG	AGGGAACAAAATGTGACCTGTAT	1q31	222	0.79
2	DIS249	TGGCATGTCTTTGAAGGAAT	TGGTTGTAGATGAGACTGGC	1q32	226	0.88
3	DIS245	GACCCCTTTACCTGAGGC	CGCTTTGATAGATTTAGCACTG	1q41	234	0.83
4	DIS229	GCTTGTTTCCATTATGGTG	ACTCTAGTTGTGTGTGAATGTATG	1q42	242	0.78
5	DIS213	CATTATCCAAGGTCAGGAGG	AGCTGTTAATCCAATCTATGATGTG	1q42	247	0.87
6	DIS225	GCCTGGGTGACAAAAGCA	TGGCCTGAATAGACCATAAAAA	1q42	248	0.80
7	DIS103	ACGAACATTCTACAAGTTAC	TTTCAGAGAACTGACCTGT	1q42	249	0.80

*†The assignment of chromosome locus and sex-averaged genetic distance is from the composite map (40). ‡Heterozygosity index.

markers in the region of mouse-human homology was tested for linkage separately, first using only data from pairs of affected siblings, then using data from all sibling pairs in the sample. Two different linkage tests were used with each sample. The advantage of an analysis that uses only affected sibling pairs is that each person in the analysis is assumed to have the disease gene of interest and other unaffected individuals who may have the gene but have not expressed it do not confound the results. In addition to these single marker analyses, we also combined the markers at the most promising region into haplotypes to maximize the information available for linkage. Given that these analyses test a specific candidate region for linkage, we selected $P < 0.05$ as our criterion to conduct subsequent analyses. Linkage at this level of significance was followed with genotyping and testing of several flanking markers to investigate the region further.

To assess linkage with the sample of clinically concordant, affected sibling pairs, we calculated the mean allele sharing identical-by-descent at the loci of interest and tested each for a significant difference from the expected value of random marker sharing, 0.5, using the SIBPAL subprogram (42) of the S.A.G.E. package (43). The test statistic follows a binomial distribution. A significant increase in allele sharing (over 0.5), is taken as evidence for linkage (44–47).

We conducted the same analyses including two additional groups of sibpairs; one in which both sibs are unaffected (clinically concordant unaffected sibpairs), and another in which one sib is affected and the other is not (clinically discordant sibpairs). At linked markers, both concordant affected and clinically concordant unaffected sibpairs are expected to demonstrate an increased sharing of marker alleles identical-by-descent, while clinically discordant pairs should exhibit a decreased sharing (45, 46). Both clinically concordant unaffected and clinically discordant sibpairs should show weaker evidence for linkage than affected sibpairs, because some unaffected siblings may carry the disease gene and yet not express it due to the lack of time or of other genetic and environmental factors which contribute to the development of disease.

With a second linkage method, we combined data from all three

groups of sibpairs using the SIBPAL subprogram (42) of the S.A.G.E. package (43) to regress the trait differences (disease status or levels of autoantibody) in sibpairs against the proportion of marker alleles that they share (48). A significantly negative regression line, representing the phenomenon that those who are phenotypically alike tend to share more alleles and those who are discordant tend to exhibit less allele sharing, is taken as evidence for linkage. If there is no linkage, this analysis yields a horizontal line with the regression coefficient not significantly different from zero. This is assessed by dividing the regression coefficient by its standard error and comparing it to a t value for the appropriate degrees of freedom.

We then constructed haplotypes of the three markers in the most promising region identified by the prior analyses and tested them simultaneously for linkage. First, we examined the observed distribution in haplotype sharing for the affected sibpairs and compared it with what is expected by chance alone using a chi-square test of goodness-of-fit test (49). A significant difference from the theoretical distribution, with the number of pairs sharing two or one alleles exceeding what is expected, is taken as evidence for linkage. Often using haplotype information will improve the significance level of a linkage result by increasing the informativeness of the markers (50). This approach is, however, vulnerable to the loss of information due to crossover events within the region of interest. We then tested the haplotype for linkage with both the mean allele sharing test and the regression test.

Results

We first performed separate linkage analyses for each of seven markers located on human chromosome 1q31–q42 region using the SIBPAL program (42) of the S.A.G.E. package (43). Except for markers *DIS510* and *DIS249* ($P > 0.1$), as shown in Table II, each of the remaining five markers provided evidence for linkage to the disease, with increased mean allele

Table II. Affected Sibpair Linkage Analysis of SLE and Chromosome 1 Markers or Haplotypes

Distance between markers	[4cM]	[8cM]	[8cM]	[5cM]	[1cM]	[< 1cM]	Haplotype
Marker (DIS) of haplotype	510	249	245	229	213	225	103
Number of informative affected sibpairs*	51	49	50	50	48	49	47
Mean allele sharing in affected sibpairs [†]	0.55	0.55	0.58	0.64	0.61	0.57	0.60
$P <$	0.11	0.10	0.02	0.0005	0.005	0.08	0.01

*The number of informative sibpairs for a given marker is reduced in case of homozygosity or allele sharing in parents. †All are greater than 0.5, the expected value for no linkage. Numbers of alleles observed range from 8 to 17 for these seven markers.

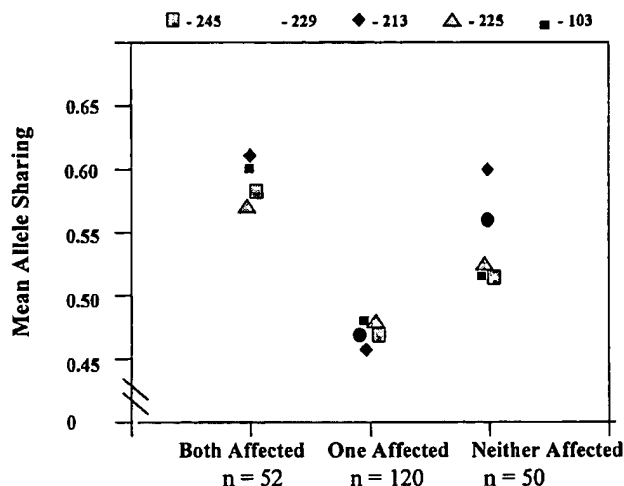


Figure 2. Mean allele sharing in SLE sibpairs for five chromosome 1 markers. Each data point represents the result for a specific marker sharing of the five separate markers. This analysis includes 52 affected sibpairs, 120 clinically discordant pairs (where one is affected and one is not), and 50 nonaffected pairs from the 43 studied SLE multiplex families.

sharing (0.57 to 0.64, $P = 0.0005$ to 0.08). The variation in levels of significance may be attributed to the informativeness of each marker and the distance of the marker from the disease gene. To examine the effect of these markers simultaneously, we chose the three contiguous markers, *DIS229*, *DIS213*, and *DIS225* (within a 7-cM region), containing the strongest linkage results and combined them into groups of linked markers on one chromosome, termed haplotypes, thus allowing us to trace their inheritance as a group. Similar to the *HLA* region, haplotypes are often more informative than individual markers. As shown in Table II, the allele sharing of this haplotype yielded similar evidence for linkage (mean allele sharing = 0.61 and $P = 0.007$). In contrast to the range of mean marker sharing in 52 affected sibpairs (0.57 to 0.64, Table II), 120 discordant pairs (where one is affected and one is not) yielded estimates of reduced marker sharing ranging between .46 and .49 for the five markers (Fig. 2). Sharing in the 50 pairs where neither was affected at the time of analysis resulted in estimates of between 0.50 and 0.60 depending on the specific marker (Fig. 2). This pattern of excess sharing at each of the loci in the candidate region for both types of concordant pairs as well as reduced sharing in clinically discordant pairs, provides further evidence for linkage. The smaller mean allele sharing for unaffected sibpairs compared to that for affected pairs (0.50–0.60 vs. 0.57–0.64) is consistent with the possibility that among the unaffected sibs, some may carry the SLE gene and will develop SLE over time, i.e., that they are actually genetically discordant.

We also conducted linear regression analyses to assess whether the pattern of differences in SLE in sibpairs varies in relation to the degree of allele sharing at the marker locus. Different disease status (one affected and one unaffected) in the sibpair should result in decreased allele sharing while the same disease status (either both affected or both unaffected) should result in increased sharing. When these differences in

Table III. Regression of Marker Sharing Identical-by-Descent and Differences in SLE Status within Pairs*

(DIS)	<i>t</i> values [†]	<i>P</i> value
245	−1.84	0.04
229	−3.41	0.0005
213	−1.87	0.03
225	−3.49	0.0004
103	−1.51	0.07
Haplotype	−1.58	0.06

*Regression analyses were based on 50 unaffected sibpairs, 120 sibpairs discordant for disease, and 52 sibpairs where both are affected. [†]Estimated regression coefficient divided by its standard error.

disease status are regressed against the degree of allele sharing for all sibpairs, a negative slope is indicative of linkage. As shown in Table III, the tested five markers as well as the haplotype showed negative slopes of the regression lines ($P = 0.0004$ to 0.07), thus further supporting the evidence for linkage in this region.

We compared our observed allele sharing in the affected pairs with a theoretical distribution for no linkage in that region (25% sharing 0 alleles, 50% sharing 1, and 25% sharing 2). The haplotype distribution defined by markers *DIS229*, *DIS213*, and *DIS225* in 50 affected sibpairs yielded a P value less than 0.01 using a χ^2 tests for goodness-of-fit against the expected distribution (Table IV). Because of crossover events within this region, two affected sibpairs were excluded from this analysis. We included the distribution delineated by ethnic groups as well; the trend of increased sharing of haplotypes was observed in affected sibpairs from these three ethnic groups individually and as a whole (Table IV). Using haplotype sharing information from the entire sample, we estimate the sibling recurrence risk ratio which may be attributable to the potential susceptibility locus in this region to be 4.17 (51).

Because this murine susceptibility region on mouse chromosome 1 was linked to the presence of IgG antichromatin antibodies in the mouse (30, 34, 35), all family members were assayed for serum IgG antibodies to chromatin. To assess whether linkage of antichromatin IgG is conserved between mice and humans, we tested serum levels of this autoantibody for linkage to this candidate region. Because levels of some autoantibodies fluctuate over time, this analysis at one time may underestimate the tested autoantibody as a quantitative trait. Nevertheless, even with this possible confounding, as shown in Table V, antichromatin IgG antibodies showed tentative

Table IV. Chromosome 1 Haplotype Sharing in Affected Sibpairs by Ethnic Group

Ethnicity	IBD Sharing		
	0	1	2
Caucasian	3	13	9
Asian	0	13	1
African American	0	8	3
Total*	3	34	13

* $\chi^2 = 10.48$, $P < 0.01$.

Table V. Regression of Marker Sharing in Sibpairs and Squared Differences of Autoantibody Levels

(DIS)	P values	
	Antichromatin	Anti-DNA
245	0.12	0.72
229	0.04	0.89
213	0.04	0.85
225	0.07	0.87
103	0.07	0.91
Haplotype	0.06	0.80

evidence for linkage ($P = 0.04$ – 0.12 , effective degrees of freedom = 70 for these five markers, and $P = 0.06$ for the haplotypes). Because IgG anti-dsDNA antibodies contribute to SLE GN, we assayed serum samples from all family members for IgG anti-dsDNA. Using the same diluted serum samples, no evidence of linkage was detected for IgG anti-DNA at any of the five markers, with P values ranging from .72 to .91. The tentative linkage result of IgG antichromatin to this region supports our mouse to human candidate region approach because this chromosome 1q region is syntenic to the murine susceptibility region containing the *Sle1*, *Lbw-7*, and *Nba2* loci, which are linked to the production of IgG antichromatin (30, 34–36). The lack of linkage of IgG anti-dsDNA to this test region is inconsistent with the recent finding that the *Nba2* locus is linked to production of autoantibodies to several nuclear antigens including dsDNA (35), but consistent with the previous report of murine linkage analyses that different chromosomal loci correlate with IgG anti-dsDNA (33).

Discussion

This study has identified a 15 cM region on chromosome 1q41–q42 that is linked to human SLE. Previously, the *MHC* region and several polymorphic gene loci have been related to human SLE by association studies (7, 10, 12–24), which may yield significant statistical results for reasons other than physical proximity to an SLE gene. We chose to test for linkage a candidate human chromosomal region syntenic to a murine susceptibility region. This murine susceptibility locus (mapped to a 67–112 cM region of chromosome 1 of NZB or NZW genetic origin) was identified repeatedly by linkage analyses of three separate crosses using two inbred strains of lupus-prone mice—(NZB × NZW) F1 and NZM/Aeg2410 mice (30, 31, 35, 36). When choosing this region for analysis, we hypothesized that (a) important susceptibility genes may be conserved from mice to humans, and (b) if susceptibility loci are conserved across species, they are likely to be conserved across some racial groups. The first hypothesis is supported by (a) the increased identity-by-descent sharing of each separately tested marker within chromosomal region 1q41–q42 (Table II), (b) the significant regression analyses relating marker sharing and disease status in all sibpairs ($P = 0.0004$ – 0.07 for each tested marker in 1q41–q42), and (c) the increased identical-by-descent sharing of haplotypes of linked markers (Table IV). Our haplotype analysis (Table IV) showing no obvious differences in sharing among the three racial groups is consistent with our second hypothesis of no racial differences in linkage in this region, although

larger sample sizes may reveal differences we cannot detect herein. Thirdly, we further hypothesized that if a specific locus is conserved across species, then the phenotypic expression of that locus could be conserved as well. The tentative linkage of IgG antichromatin to markers of this region is supportive evidence that the phenotypic expression of the locus may be conserved between mice and humans.

Among these seven markers tested, *DIS229* appears to be the most promising marker linked to both the disease and to the quantitative trait of IgG antichromatin antibodies. This might be attributed to the informativeness of this marker and/or the distance of this marker from a lupus putative susceptibility gene. Because the heterogeneity index of *DIS229* is similar if not lower than the other tested markers, it is tempting to speculate that a disease causing gene may be nearest to *DIS229* among the markers we have tested. A previous linkage report in human SLE observed an optimal lod score of 2.38 of a marker at 1q23 in a small sample of four African-American families (25).

Our observation of a conserved susceptibility region between mouse and human SLE is not a common finding in autoimmune diseases. For example, linkage analyses of human type I insulin-dependent diabetes mellitus (IDDM) sibpairs and diabetes in the NOD mouse showed that most susceptibility loci did not correlate well with two exceptions: (a) IDDM7 maps to the syntenic region of *Idd5*, and (b) IDDM1/*Idd1* are linked to the MHC region in both species (8, 52, 53). This lack of correlation between mouse and human for most diabetogenic loci may result from the fact that the NOD mice is the only mouse model for spontaneous IDDM, whereas there are several inbred strains of lupus-prone mice. Our candidate chromosomal region is syntenic to overlaps of murine susceptibility loci derived from more than one genetic origin. If susceptibility genes are conserved across different mouse loci in several strains, they are more likely to be conserved across species. This may account for the observed linkage to the disease and to the IgG antichromatin phenotype which appears to be conserved between mouse and human as well. Recent success of mapping a putative susceptibility locus for multiple sclerosis in the 5p14–p12 region, which is syntenic to the murine locus *Eae2*, further supports the utility of this mouse to human approach (54).

Within the candidate chromosomal region 1q21–q42, *FcyRIIIa* alleles have been associated with SLE GN in African-Americans but not in Afro-Caribbean or Chinese (22, 55). The role of *FcyRIIIa* alleles in Caucasian SLE patients is controversial; an association with SLE GN has been found in the Dutch (23) but not in the American or British Caucasian cohorts (22, 55). The distance (> 60 cM) between the 1q41–q42 region and the *FcyRIIIa* gene (1q23) makes it unlikely that this gene accounts for our observed linkage. The *CRI* gene (complement receptor one, previously C3b/C4b receptor) maps to chromosome 1q32. A polymorphism for low expression of *CRI* has been suggested to be a risk factor for SLE (17), although later studies suggested low expression was acquired (56). The increased marker sharing at the 1q41–q42 region but not at the 1q31–q32 region makes the *CRI* gene unlikely to account for our results. Therefore, this chromosome 1q41–q42 region may contain a currently unidentified disease-causing gene or genes. Our mapped interval currently spans 15 cM and may contain up to 500 genes. Several genes (*TGFB2*, *HLXI*, and *ADPRT*) located within this region can encode proteins which may play a

role in development of SLE (57–59). *TGFB2* (transforming growth factor beta-2) can suppress IL-2 dependent T cell growth (60) and administration of a *TGFB* cDNA expression vector to MRL/lpr mice decreases serum IgG antichromatin and delays the onset of SLE (61). *HLX1* (homeo box 1) is expressed in hematopoietic progenitors and activated lymphocytes and may regulate development of CD4⁺ T cells (62). *ADPRT* (ADP-ribosyltransferase) is induced by DNA damage and plays a role in cellular repair (63). The report that SLE patients and their family members had decreased poly(ADP-ribose) metabolism makes *ADPRT* a possible candidate (64). Further analyses of this candidate region (1q41–q42) are in progress and should lead to eventual identification of a specific susceptibility gene or genes for human SLE.

In summary, the data presented herein provide support for the hypothesis that a genetic region homologous to a portion of chromosome 1 in the mouse provides genetic susceptibility to SLE in species as widely divergent as man and mouse. This locus appears to predispose to SLE in all ethnic groups examined.

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